

ALKALOIDS OF *DEHAASIA TRIANDRA**

SHENG-TEH LU, IAN-LIH TSAI and SHIOW-PIAW LEOU

School of Pharmacy, Kaohsiung Medical College, 100 Shih-Chuan First Road, San-Min District, Kaohsiung, Taiwan, R.O.C.

(Received 16 February 1988)

Key Word Index—*Dehaasia triandra*; Lauraceae; Iau-Guoo-Nan; bisbenzylisoquinoline alkaloids; dehatridine; dehatrine.

Abstract—Separation of the basic fraction from *Dehaasia triandra* afforded two new bisbenzylisoquinoline alkaloids, dehatridine and dehatrine, along with six known alkaloids, isocorydine, corytuberine, atheroline, nantenine, obaberine and a quaternary aporphine alkaloid, xanthoplanine (5).

INTRODUCTION

The Formosan lauraceous plant, *Dehaasia triandra* [*Endiandra lanyuensis* Chang; *D. lanyuensis* (Chang) Kostermans] (Chinese name; Iau-Guoo-Nan) [1] is a small evergreen tree growing on Orchid Island and in the Philippines. Detailed research on the alkaloids of *Dehaasia* sp. is still rare, although preliminary investigations showed that these plants contain alkaloids [2–4]. In a previous paper [5], two alkaloids, isocorydine (1) an aporphine, and obaberine (6) bisbenzyltetrahydroisoquinolines, were obtained. Because there are still several alkaloids which are not yet identified, a further study was made for confirmation of these unknown compounds. As a result two new bisbenzylisoquinoline alkaloids, dehatridine (7) and dehatrine (8), along with six known alkaloids, isocorydine (1), corytuberine (2), atheroline (3), nantenine (4), obaberine (6) and xanthoplanine (5) were isolated. Except for 6 and 8 which were obtained from the trunk, the other six alkaloids were obtained from the leaves. This paper reports the isolation and characterization of these alkaloidal components from *D. triandra*.

RESULTS AND DISCUSSION

Extraction and separation of alkaloids were performed by the usual procedures as described in the Experimental. The following alkaloids were isolated: (a) aporphines: (+)-isocorydine (1), (+)-corytuberine (2), (+)-nantenine (4) and (+)-xanthoplanine (5); (b) oxoaporphines: atheroline (3); (c) bisbenzylisoquinolines: obaberine (6) dehatridine (7) and dehatrine (8) [Table 1].

Isocorydine (1) was identified by direct comparison [mmp, TLC, UV, IR (KBr), ¹H NMR] with an authentic sample. Corytuberine (2) was derived from corytuberine (2) MeI which was identified by comparison of mmp and IR (KBr) with authentic magnoflorine (9) I. The xanthoplanine (5) iodide was recognized as *N*-methyl laurotetanine (10) MeI by comparison of mmp, IR (Nujol) and UV.

The physical data [mp, [α]_D, UV, IR (KBr) and ¹H NMR] of nantenine (4) were in agreement with those of nantenine (4) [6–8]. With Ac₂O in pyridine, atheroline (3) gave an *O*-acetyl derivative as yellow needles, mp 190–193°, which was identified by comparison with authentic *O*-acetylatheroline (12) [9, 10] by mmp, TLC and IR (KBr). The HBr salt of obaberine (6), a bisbenzyltetrahydroisoquinoline alkaloid, was identified [mmp, TLC, IR (KBr)] by comparison with an authentic sample.

Dehatridine (7) was isolated as colourless needles, mp 274–276° (MeOH), and [α]_D²³ +98° (MeOH, *c* 0.1). The molecular formula C₃₅H₃₂N₂O₆ belonging to bisbenzylisoquinolines was determined by elemental analysis and mass spectrometry [*m/z* 576 [M]⁺]. UV indicated that there is a benzylisoquinoline moiety and a phenolic OH group in the molecule [$\lambda_{\max}^{\text{EtOH}}$ nm: 202, 243, 282 and 335, shows bathochromic shift on addition of KOH]. In the mass spectrum, there are the intensive fragments at *m/z* (%) 576 [M]⁺, 100, 575 [M–1]⁺, 92, 190 (30) and 174 (60). This fragmentation indicates that an aromatic isoquinoline moiety is present in the molecule [11, 12]. In the ¹H NMR, there are two mutually coupled signals at δ 7.43 and 8.20 (each 1H, *d*, *J* = 5.48 Hz) corresponding to the aromatic protons existing on a substituted pyridine ring. On the other hand, there is a one pair doublet with a larger coupling constants at δ 3.85 and 4.66 (each 1H, *d*, *J*_{gem} = 12.5 Hz) corresponding to the benzylic methylene protons adjacent to the pyridine ring. Moreover, there is one *N*-Me at δ 2.56 (3H, *s*), two OMe at δ 3.38 and 3.82 (each 3H, *s*) and two phenolic OH groups at δ 8.65 and 9.03 (each 1H, *s*) which were exchangeable with D₂O. On the one hand, *O*-methylation with diazomethane on dehatridine (7) gave *O*,*O*-dimethyldehatridine (13). In the aromatic proton regions, there are 10 aromatic protons at δ 5.92, 6.62, 7.02 (each 1H, *s*), 6.60 (1H, *d*, *J*_m = 2 Hz), 6.69 (1H, *d*, *J*_o = 8.2 Hz), 6.82, 6.84, 6.94, 7.19, 7.66 (each 1H, *dd*, *J*_o = 8.2 Hz, *J*_m = 2 Hz). From their chemical shifts, each of them was assigned to H-8', H-5', H-5, H-10, H-13, H-10', H-14, H-11', H-13' and H-14', respectively. Spin decoupling supported these assignments. Although there is a benzylisoquinoline moiety in the molecule, reduction with NaBH₄, Zn+HOAc, Zn/Hg, Pd/C+H₂, Na or Na/Hg on dehatridine (7) did not give the expected

*Part 31 in the series 'Studies on the Alkaloids of Formosan Lauraceous Plants'. For Part 30, See Lu, S.-T and Tsai, I.-L. (1988) *Heterocycles* (in press).

Table 1. Alkaloids isolated from *Dehaasia triandra*

Bases	mp (°)	Leaves (6.19 kg)		Wood (12.64 kg)	
		yields g	%	yields g	%
A. Tertiary phenolic:					
Isocorydine(1)	187–188	4.700	0.0680	---	---
Corytuberine (2)	239–242	0.070	0.0011	---	---
Atheroline (3)	250–252	0.026	0.0004	---	---
Dehatridine (7)	274–276	0.950	0.0153	---	---
B. Quaternary:					
Xanthoplanine (5)	Cl salt	---	---	---	---
	218–220				
	picrate	7.450	0.1200	---	---
	227–228				
C. Tertiary non-phenolic:					
Nantenine (4)	131–134	0.012	0.0002	---	---
Obaberine (6)	HBr salt	---	---	8.42	0.067
	263–265				
	(decomp.)				
Dehatrine (8)	158–160	---	---	0.348	0.003

product, tetrahydrodehatridine [11]. Sodium/liquid ammonia cleavage of *O,O*-diethyldehatridine (14) afforded (+)-*N*-methylcoclaurine (15) from the phenolic base part and (±)-*O,O*-diethylcoclaurine (16) from the non-phenolic base part. It was assumed that 7,4'-diethoxy-6-methoxybenzylisoquinoline (16a) produced by Na/liq. NH₃ cleavage of the ether linkages of *O,O*-diethyldehatridine (14) was reduced to (±)-*O,O*-diethylcoclaurine (16). Both (+)-*N*-methylcoclaurine (15) and (±)-*O,O*-diethylcoclaurine (16) were identified [TLC, IR (KBr), ¹H NMR] by comparison with authentic samples derived from (±)-coclaurine (18), respectively. Therefore, the two ether linkages between the benzyltetrahydroisoquinoline moiety and the benzylisoquinoline moiety of dehatridine (7) are assumed to be at 8-7', 11-12' as found in the berbamine series. Moreover, the absolute configuration of H-1' is *S*.

Dehatrine (8) was isolated as colourless needles, mp 158–160° (EtOH) and $[\alpha]_D^{27} + 27^\circ$ (CHCl₃, *c* 1.0) from the tertiary non-phenolic base part. It is a bisbenzylisoquinoline alkaloid [C₃₁H₃₈N₂O₆, *m/z* 606 [M]⁺] whose UV and IR suggested that there is an imine group in the molecule [$\lambda_{\max}^{\text{EtOH}}$ nm: 223, 281, 310; ν_{\max}^{KBr} cm⁻¹: 1620 (C=N)]. The ¹H NMR displayed a methylene group adjacent to the C=N of a 3,4-dihydroisoquinoline moiety at δ 4.12 and 5.32 (each 1H, *d*, *J*_{gem} = 2 Hz), one N-Me at δ 2.46 (3H, *s*), and four OMe at δ 3.51, 3.70 (each 3H, *s*) and 3.91 (6H, *s*). In the aromatic proton regions, there are 10 aromatic protons at 6.05, 6.42, 6.63 (each 1H, *s*, H-8',5',5), 6.66–6.88 (3H, *m*, H-10, 13, 14), 7.01 (2H, *d*, *J* = 8 Hz, H-11',13'), and 7.35 (2H, *d*, *J* = 8 Hz, H-10',14'). Sodium/liquid ammonia cleavage reaction afforded *R*-(−)-*O*-methylarmepavine ($[\alpha]_D^{29} - 107^\circ$) (17) and (±)-coclaurine ($[\alpha]_D^{29} \pm 0^\circ$) (18) respectively, which were identified [mp, TLC, IR (KBr), ¹H NMR] by comparison with authentic samples. Reduction with NaBH₄ of dehatrine (8) afforded 1,2-dihydrodehatrine (19), a bisbenzyltetrahydroisoquinoline alkaloid, [$\lambda_{\max}^{\text{EtOH}}$ 228,

280 nm ν_{\max}^{KBr} absent 1620 cm⁻¹ peak, the signals at δ 4.12 and 5.32 disappeared]. [13]. *N*-Methylation of 1,2-dihydrodehatrine (19) gave a mixture of isotetrandrine (20) and phaeanthine (21) which were separated by prep. TLC. The former was identified [mp, TLC, IR (KBr), ¹H NMR] by comparison with authentic isotetrandrine (20). The physical data (mp, $[\alpha]_D$, UV) of the latter was compared with those of phaeanthine (21) [13]; they were all in agreement. Therefore, the two ether linkages between the benzyltetrahydroisoquinoline moiety and the 3,4-dihydrobenzylisoquinoline moiety of dehatrine (8), are assumed to be at 8-7', 11-12' as in the berbamine series. The absolute configuration of H-1 is *R*.

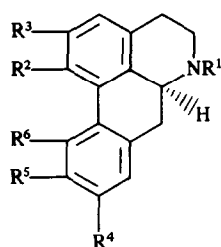
EXPERIMENTAL

Mps: uncorr. ¹H NMR were recorded at 60, 100 or 270 MHz with TMS as int. std; chemical shifts are recorded in δ (ppm) units. MS were measured at 75 eV. Silica gel (60–230 mesh) (Merck) and neutral alumina (Merck) were used for CC and silica gel GF-254 for TLC.

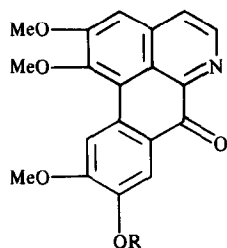
Leaves and trunks of *D. triandra* Merr. were collected at Orchid Island, in August, 1976. Air-dried plant material, was extracted with warm EtOH and the EtOH soln concd under red press to leave a dark brownish viscous residuc. Bases in the EtOH extracts were extracted with 3% HOAc and the acidic soln treated with alkalis and CHCl₃-Et₂O.

Isocorydine (1). Part B (8.7 g from leaf extract) crystallized in contact with Me₂CO as colourless prisms, mp 187–188° (lit. 185–186°) and $[\alpha]_D^{23} + 205^\circ$ (MeOH, *c* 0.2) (lit. 194°) [14]. Gibbs test positive. UV and ¹H NMR were the same as ref. [5]. Identified [mp, TLC, IR (KBr)] by comparison with authentic material.

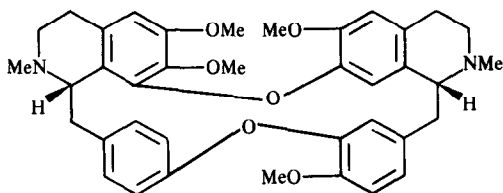
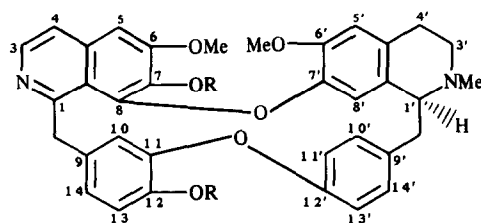
Corytuberine (2). The residue remaining after the separation of isocorydine (1) was placed on a silica gel column and eluted with CHCl₃ gradually enriched with Me₂CO. Fractions eluting with CHCl₃-Me₂CO (1:1) provided corytuberine (2) as colourless prisms, mp 239–242° (lit. 240°) [15], $[\alpha]_D^{23} + 276^\circ$ (MeOH, *c* 0.1)



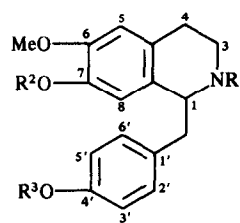
	R ¹	R ²	R ³	R ⁴	R ⁵	R ⁶
1	Me	OMe	OMe	H	OMe	OH
2	Me	OH	OMe	H	OMe	OH
4	Me	OMe	OMe	OCH ₂ O	H	
5	(Me) ₂	OMe	OMe	OH	OMe	H
9	(Me) ₂	OH	OMe	H	OMe	OH
10	Me	OMe	OMe	OH	OMe	H
11	(Me) ₂	OMe	OMe	OMe	OMe	H



- 3** R = H
12 R = Ac

**6**

- 7** R = H
13 R = Me
14 R = Et



- 15** R¹ = Me, R² = R³ = H
16 R¹ = H, R² = R³ = Et
17 R¹ = R² = R³ = Me
18 R¹ = R² = R³ = H

(lit. 286°) [16]. MS m/z : 327 ($[M]^+$). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400 (OH). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 222 (4.4), 268 (4.0), 306 (3.5). UV $\lambda_{\text{max}}^{\text{EtOH} + \text{KOH}}$ nm (log ϵ): 208 (4.5), 226 (4.4), 276 (3.7), 318 (3.7). ^1H NMR (100 MHz, DMSO- d_6): δ 2.55 (3H, s, NMe), 3.75 (6H, s, OMe \times 2), 4.50 (2H, br s, OH \times 2, D₂O exchangeable), 6.53 (1H, s, H-3), 6.58 (1H, d, J = 8 Hz, H-9), 6.80 (1H, d, J = 8 Hz, H-8). MeI (**9**): colourless needles, mp 245–248°; $[\alpha]_D^{23} + 195^\circ$ (MeOH, c 0.1). Identified [mmp, TLC, IR (KBr)] by comparison with magnoflorine (**9**) I.

Atheroline (3). Part E from the extract (65 g) of the leaves was chromatographed on silica gel and eluted with CHCl_3 gradually enriched with Me_2CO . The fractions eluting with CHCl_3 – Me_2CO (2:1) afforded atheroline (**3**) as reddish needles (MeOH), mp 250–252° (lit. 250–260°) [9]. $[\alpha]_D^{23} \pm 0^\circ$ (MeOH, c 0.1). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 242 (4.6), 272 (4.6), 292 (4.4), 352 (4.1), 380 (4.0), 433 (3.7), $\lambda_{\text{max}}^{\text{EtOH} + \text{KOH}}$ nm (log ϵ): 252 (4.7) 299 (4.6), 323 (4.6), 400 (4.2). $\text{C}_{19}\text{H}_{15}\text{O}_5\text{N}$, MS m/z (%): 337 ($[M]^+$, 100), 322 (30), 307 (22), 294 (26.8), 279 (17), 262 (19.5). ^1H NMR (100 MHz, DMSO- d_6): δ 4.00 (6H, s, OMe \times 2), 4.05 (3H, s, OMe), 7.60 (1H, s, H-3), 7.73 (1H, s, H-8), 8.03 (1H, d, J = 5 Hz, H-4), 8.70 (1H, s, H-11), 8.80 (1H, d, J = 5 Hz, H-5). Atheroline (**3**) treated with Ac_2O and pyridine gave *O*-acetylatheroline (**12**) as light yellowish needles (Me_2CO), mp 190–193° (lit. 190–195°) [9]. It was

identified [mmp, TLC, IR (KBr)] by comparison with authentic material.

Nantenine (4). Part G (4.8 g) from extract of leaves was placed on an alumina column and eluted with CHCl_3 to provide nantenine (**3**) as colourless needles (Me_2CO), mp 131–134° (lit. 139–141°) [6] and $[\alpha]_D^{20} + 79^\circ$ (MeOH, c 0.1) (lit. +93°) [7]. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) [8]. 223 (4.56), 272 sh (3.9), 282 (4.0), 308 (4.1), 318 sh (4.0). ^1H NMR (60 MHz, CDCl_3): δ 2.50 (3H, s, NMe), 3.63, 3.83 (each 3H, s, OMe), 5.93 (2H, s, OCH_2O), 6.57 (1H, s, H-3), 6.73 (1H, s, H-8), 7.90 (1H, s, H-11) [7]. The above data correspond with those of nantenine [6–8].

Xanthoplanine (5). When the quaternary base part of the leaves was purified by the ordinary method through the base Reineckate \rightarrow base sulphate \rightarrow base chloride, xanthoplanine (**5**) was obtained as the Cl salt [colourless needles, mp 218–220° (lit. 218–220°) [18], $[\alpha]_D^{18} + 85^\circ$ (MeOH, c 0.1); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 225 (4.0), 286 (3.6), 310 (3.7); $\lambda_{\text{max}}^{\text{MeOH} + \text{KOH}}$ nm (log ϵ): 214 (4.5), 254sh (3.8), 343 (4.1); ^1H NMR (60 MHz, TFA): δ 3.20, 3.50 (each 3H, s, $=\text{N}^+\text{Me}_2$), 3.88 (3H, s, OMe), 4.06 (6H, s, OMe \times 2), 6.90, 7.03, 8.07 (each 1H, s, H-3, 8, 11). Iodide: colourless prisms, mp 190–191° (lit. 148–149°) [18]. Identified [mmp, UV, IR (KBr)] with authentic xanthoplanine I. Picrate: yellow prisms, mp 227–228° (Me_2CO) (lit. 228° decomp.) [19]. *O*-Methylxanthopla-

(+)-N-Methylcoclaurine (15). $[\alpha]_D^{23} + 64^\circ$ (MeOH, *c* 0.1). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 231 (4.1) and 288 (3.6). UV $\lambda_{\text{max}}^{\text{MeOH} + \text{KOH}}$ nm (log ϵ): 223 (4.3), 250 (4.0), 296 (3.6). $^1\text{H NMR}$ (60 MHz, CDCl_3): δ 2.47 (3H, s, NMe), 3.83 (3H, s, OMe), 5.03 (2H, s, OH \times 2, D_2O exchangeable), 6.37 (1H, s, H-8), 6.53 (1H, s, H-5), 6.63 (2H, *d*, $J_o = 8$ Hz, H-3', 5'), 6.93 (2H, *d*, $J_o = 8$ Hz, H-2', 6'). Identified [TLC, UV, $^1\text{H NMR}$, IR (CHCl_3)] by comparison with an authentic sample.

Dehatrine (8). The Me_2CO soln after separating obaberine (6) HBr was evapd under red. pres. at room temp to leave a dark brownish viscous residue. The residue was dissolved in CHCl_3 and the soln extracted with 2% H_2SO_4 . The H_2SO_4 soln was basified with NH_4OH and extracted with Et_2O . The Et_2O soln was dried (K_2CO_3) and evapd to leave a yellowish viscous residue. This was placed on a silica gel column and eluted with CHCl_3 to give dehatrine (8) as colourless needles (EtOH), mp 158–160° and $[\alpha]_D^{27} + 27^\circ$ (CHCl_3 , *c* 1.0). MS *m/z* (%): 606 ($[\text{M}]^+$, 100), 605 (70), 591 (35), 303 (68), 280 (15), 204 (20), 155 (15), 141 (45). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 223 (4.5), 281 (3.8), 310 (3.5). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 1620 (C=O). $^1\text{H NMR}$ (100 MHz, CDCl_3): δ 2.46 (3H, s, NMe), 3.51 (3H, s, OMe), 3.70 (3H, s, OMe), 3.91 (6H, s, OMe \times 2), 4.12 and 5.32 [each 1H, *d*, $J = 2$ Hz, (benzylic methylene)], 6.05 (1H, s, H-8'), 6.42 (1H, s, H-5'), 6.63 (1H, s, H-5), 6.66–6.88 (3H, *m*, Ar. H), 7.01 (2H, *d*, $J = 8$ Hz, H-11', 13'), 7.35 (2H, *d*, $J = 8$ Hz, H-10', 14'). Anal.: calcd for $\text{C}_{37}\text{H}_{38}\text{O}_6\text{N}_2 \cdot \text{H}_2\text{O}$, C, 70.25; H, 6.11; N, 4.34. Found: C, 70.41; H, 6.11; N, 4.28.

Na/liq. NH_3 cleavage of dehatrine (8). When a THF soln of 8 (450 mg) was treated with Na (700 mg)/liq. NH_3 (150 ml) as described for *O,O*-diethyldehatridine (14), *R*-(–)-*O*-methylarmepavine (17) (230 mg) and (±)-coclaurine (18) (205 mg) were obtained.

R-(–)-*O*-Methylarmepavine (17). Oxalate: colourless needles, mp 170–175° (EtOH) and $[\alpha]_D^{29} - 107^\circ$ (MeOH, *c* 1.25). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 212 (4.0), 225 (4.0), 282 (3.5). MS *m/z* (%): 327 ($[\text{M}]^+$, 2), 206 (100), 191 (54), 190 (35), 152 (53), 151 (33). Anal.: calcd for $\text{C}_{20}\text{H}_{25}\text{NO}_3 (\text{COOH})_2 \cdot \text{H}_2\text{O}$, C, 60.68; H, 6.71; N, 3.22. Found: C, 60.83; H, 6.53; N, 3.01. Identified [mmp, UV, IR (KBr)] by comparison with an authentic sample. $^1\text{H NMR}$ (100 MHz, CDCl_3) of *R*-(–)-*O*-methylarmepavine (17) generated from oxalate: δ 2.52 (3H, s, NMe), 3.56, 3.76 and 3.82 (each 3H, s, OMe), 6.00 (1H, s, H-8), 6.52 (1H, s, H-5), 6.78 (2H, *d*, $J_o = 8$ Hz, H-3', 5'), 7.00 (2H, *d*, $J_o = 2$ Hz, H-2', 6'). MeI: colourless prisms, mp 129–130° (swelling) (MeOH). Anal.: calcd. for $\text{C}_{18}\text{H}_{28}\text{O}_3\text{NI} \cdot 1/2 \text{H}_2\text{O}$, C, 50.81; H, 6.29; N, 2.80. Found: C, 50.56; H, 6.32; N, 2.82. Identified [mmp, IR (KBr)] by comparison with an authentic sample.

(±)-Coclaurine (18). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 207 (4.0), 225 (3.7), 283 (3.3). MS *m/z* (%): 285 ($[\text{M}]^+$, 4), 178 (67), 177 (100), 162 (79), 161 (31), 148 (21). Anal. Calcd. for $\text{C}_{17}\text{H}_{19}\text{NO}_3 \cdot \text{HBr} \cdot 3/4 \text{H}_2\text{O}$: C, 53.76; H, 5.71; N, 3.69. Found: C, 53.73; H, 5.81; N, 3.77. $^1\text{H NMR}$ (100 MHz, CDCl_3): δ 3.84 (3H, s, OMe), 4.08 (2H, *br* s, OH \times 2, D_2O exchangeable), 6.52 (1H, s, H-8), 6.60 (1H, s, H-5), 6.72 (2H, *d*, $J = 8$ Hz, H-3', 5'), 7.00 (2H, *d*, $J = 8$ Hz, H-2', 6'). HBr: Light orange needles, mp 255–256° (decomp.) (MeOH). $[\alpha]_D^{30} \pm 0^\circ$ (MeOH, *c* 0.1). Identified [mmp, IR (KBr)] by comparison with an authentic sample.

(±)-1',2'-Dihydrodehatrine (19). Dehatrine (8) (60 mg) was reduced by NaBH_4 in MeOH at room temp to afford (±)-1',2'-dihydrodehatrine (19) (58 mg). $[\alpha]_D^{23} - 52^\circ$ (MeOH, *c* 0.1). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 228 (4.6), 280 (3.6). $^1\text{H NMR}$ (60 MHz, CDCl_3): δ 2.30 (3H, s, NMe), 3.20, 3.63, 3.73 and 3.90 (each 3H, s, OMe), 6.00 (1H, s, H-8'), 6.27 (1H, s, H-5'), 6.50 (1H, s, H-5), 6.63–7.25 (7H, *m*, Ar. H).

N-Methylation of 1', 2'-dihydrodehatrine (19). Ten drops of 35% HCHO were added to a MeOH soln of 19 (58 mg) and stirred for 10 min at room temp. NaBH_4 (1 g) divided into three

parts was then added to the MeOH soln while stirring. After 2 hr, the mixt. was acidified with 10% HOAc and evapd to remove MeOH under red. pres. The conc soln was basified with NH_4OH and extracted with Et_2O . The Et_2O soln was dried (K_2CO_3) and evapd to leave a light yellowish viscous residue. The residue $[\alpha]_D^{23} - 42^\circ$ (MeOH, *c* 0.1) was divided into two parts (R_f 0.56 and 0.6) by prep. TLC [silica gel, CHCl_3 –MeOH (13:1)]. Isotetrandrine (20) (30 mg) and phaeanthine (21) (20 mg) were obtained from the zone R_f 0.56 and R_f 0.6, respectively.*

Isotetrandrine (20). Colourless prisms (Me_2CO), mp 189–190.5° and $[\alpha]_D^{23} + 140^\circ$ (MeOH, *c* 0.1). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 216 (4.9), 240 sh (4.3), 284 (3.8). $^1\text{H NMR}$ (60 MHz, CDCl_3): δ 2.23, 2.53 (each 3H, s, NMe), 3.60, 3.73, 3.90 (each 3H, s, OMe), 6.00 (1H, s, H-8'), 6.30 (1H, s, H-5'), 6.40 (1H, s, H-5), 6.50–7.20 (7H, *m*, Ar. H). Identified [mmp, TLC, IR (KBr), $^1\text{H NMR}$] by comparison with an authentic sample.

Phaeanthine (21). Colourless needles (Me_2CO), mp 142–145° and $[\alpha]_D^{23} - 230^\circ$ (MeOH, *c* 0.1). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 218 (4.5), 240 sh (4.2), 280 (4.1). $^1\text{H NMR}$ (60 MHz, CDCl_3): δ 2.33, 2.60 (each 3H, s, NMe), 3.17, 3.33, 3.70 and 3.90 (each 3H, s, OMe), 5.97 (1H, s, H-8'), 6.23 (1H, s, H-5'), 6.47 (1H, s, H-5), 6.53 (1H, s, H-10), 6.73–7.40 (6H, *m*, Ar. H). Its mp, $[\alpha]_D$, UV, MS and $^1\text{H NMR}$ were in agreement with those of phaeanthine in the lit. [13].

Acknowledgements—We are very grateful to Dr H. Ishii (Chiba University) for assistance with measurements of $^1\text{H NMR}$ and spin decoupling, to Dr Tian-Shung Wu (Providence College of Arts and Science) for assistance with MS and elemental analysis, to Dr G. K. Douglas (Tasmania University) for an authentic sample of *O*-acetylatheroline and to Prof. Ih-Sheng Chen (Kaohsiung Medical College) for assistance with collection of the plant material.

REFERENCES

1. Chang, C. E. (1963) *Bot. Bull. Acad. Sinica* **4**, 60.
2. Greshoff, M. (1980) *Bereich* **23**, 3546.
3. Henry, T. A. (1949) *The Plant Alkaloids* 4th Edn, p. 319. J & A Churchill, London.
4. U. S. Department of Agriculture (1961) *Alkaloid Bearing Plants and Their Contained Alkaloids*, Technical Bulletin No. 1234, p. 97.
5. Lu, S. T. and Wang, E. C. (1977) *J. Taiwan Pharm. Assoc.* **29**, 49.
6. Johns, S. R., Lamberton, J. A. and Sioumis, A. A. (1967) *Aust. J. Chem.* **20**, 1457.
7. Johns, S. R., Lamberton, J. A. and Sioumis, A. A. (1966) *Aust. J. Chem.* **19**, 2331.
8. Craig, J. C. and Roy, S. K. (1965) *Tetrahedron* **21**, 385.
9. Bick, I. R. C. and Douglas, G. K. (1965) *Tetrahedron Letters* 2399.
10. Bick, I. R. C. and Douglas, G. K. (1965) *Tetrahedron Letters* 4655.
11. Patra, A., Freyer, A. J., Guinaudeau, H., Shamma, M., Tantisewice, B. and Pharadai, K. (1986) *J. Nat. Prod.* **49**, 424.
12. Lavault, M., Fournet, A., Guinaudeau, H. and Bruneton, J. (1986) *Chem. Pharm. Bull.* **34**, 1148.
13. Inubushi, Y., Masaki, Y., Matsumoto, S. and Takami, F. (1969) *J. Chem. Soc. (C)* 1547.

*Inubushi *et al.* have separated these two compounds as the corresponding picrates [13].

14. Johns, S. R. and Lomberton, J. A. (1967) *Aust. J. Chem.* **20**, 1277.
15. Gadamer, T. (1911) *Arch. Pharm. (Weinheim)* **249**, 641.
16. Nijland, M. M. (1965) *Pharm. Weekbl.* **100**, 88.
17. Bick, I. R. C., Bovie, J. H. and Douglas, G. K. (1967) *Aust. J. Chem.* **20**, 1403.
18. Ishii, H. (1961) *Yakugaku Zashii* **81**, 243.
19. Ishii, H. and Harada, K. (1961) *Yakugaku Zashii* **81**, 238.